



The Open Microbiology Journal

Content list available at: www.benthamopen.com/TOMICROJ/

DOI: 10.2174/1874285801610010057



Isolation, Detection, and Characterization of Enterotoxigenic *Bacteroides fragilis* in Clinical Samples

Payam Fathi and Shaoguang Wu*

Department of Medicine, Division of Infectious Disease, Johns Hopkins University, Baltimore, MD, USA

Abstract: *Bacteroides fragilis* is an extensively studied anaerobic bacterium comprising the normal flora of the human gut. *B. fragilis* is known to be one of the most commonly isolated species from clinical samples and has been shown to cause a wide range of pathologies in humans [1, 2]. As an opportunistic pathogen *B. fragilis* can cause abscess formation and bacteremia [2]. Additionally in its enterotoxigenic form, *B. fragilis* is a known cause of diarrheal illness, is associated with inflammatory bowel disease, and has been recently characterized in patients with colon cancer [3 - 5]. As research in the field of the gut microbiome continues to expand at an ever increasing rate due to advances in the availability of next generation sequencing and analysis tools it is important to outline various molecular methods that can be employed in quickly detecting and isolating relevant strains of *B. fragilis*. This review outlines methods that are routinely employed in the isolation and detection of *B. fragilis*, with an emphasis on characterizing enterotoxigenic *B. fragilis* (ETBF) strains.

Keywords: *Bacteroides fragilis*, diarrheal illness, enterotoxigenic form, human gut, non-enterotoxigenic.

INTRODUCTION

B. fragilis and ETBF

B. fragilis is a gram-negative, bile resistant, rod shaped member of the *Bacteroidetes* phylum and *Bacteroides* genus. It is referred to as a member of the *B. fragilis* group, which is used to describe species of *Bacteroides* based on phylogenetic similarities [6 - 8]. *B. fragilis* is described as being present in two different variations: Non-enterotoxigenic (NTBF) and Enterotoxigenic (ETBF) [7, 9]. Non-enterotoxigenic *B. fragilis* are characterized as symbiotic opportunistic pathogens assisting with polysaccharide metabolism and activating protective host immune responses [10]. The enterotoxigenic form of *B. fragilis* (ETBF) results from *B. fragilis* strains which contain the *B. fragilis* toxin (BFT) encoded by a gene (*bft*) located in a 6 kb pathogenicity island termed BfPAI along with an 18 kb flanking DNA region [11]. BFT, also called fragilysin, is a zinc-metalloprotease transcribed as a 397 amino acid protein holotoxin with an 18 amino acid signal sequence, a 193 amino acid propeptide sequence, and a 186 toxin peptide sequence. Following cleavage of the signal and propeptide domains, biologically active BFT is 20 kDa in size [12]. The toxin has been shown to be present in three different isotypes with varying toxicity: BFT-1, BFT-2, and BFT-3. BFT-2 is more toxic than BFT-1 or BFT-3 [13, 14]. BFT induces toxin activity by binding to a specific, as of yet uncharacterized, receptor on the surface of intestinal epithelial cells [15]. Following binding BFT results in the rapid cleavage of the E-cadherin protein in an ATP dependent fashion resulting in decreased barrier function and subsequent permeability of the epithelial cell barrier [16]. Downstream effects of the cleavage of E-cadherin include the induction of β -catenin signaling pathways which causes an increase in intestinal epithelial cell proliferation. Furthermore BFT toxin binding has been shown to also activate NF- κ B signaling pathways [14, 17].

* Address correspondence to this author at the Department of Medicine, Division of Infectious Disease, Johns Hopkins University School of Medicine, Baltimore, MD, 21231, USA; E-mail: sgwu@jhmi.edu

Role in Gut Dysbiosis

B. fragilis was initially implicated as a causative agent in lamb diarrheal diseases in the 1980s [18]. Further studies have implicated ETBF in diarrheal disease in humans and other animal species [5, 19]. ETBF infections are now known to cause inflammatory diarrheal illnesses in children and adults [3]. The implication that ETBF is a cause of inflammatory disease is further supported by experiments in animal models which have shown that ETBF infection in mice cause acute symptomatic colitis and persistent inflammation of the colon while NTBF colonized mice exhibited histopathological normality [20]. The ability of ETBF to cause a long lasting inflammatory response in animal models contributes to its potential role as a causative agent in colon cancer. Murine studies utilizing APC/Min mice show that ETBF gavage results in Stat3 and T_H-17 induced colitis which leads to the development of colon tumors in these mice [21, 22]. Studies conducted in human colon cancer populations have shown an increased rate of detection of the *bft* gene in *B. fragilis* isolates of stool samples from patients with colorectal cancer when compared to control populations [23]. More recently, an examination of the mucosa isolated from tumor samples of colorectal cancer patients show higher levels of *bft* in the colon tumor mucosa compared to paired normal tissues [4]. The ability to properly isolate and characterize *B. fragilis* strains in clinical samples is important in further understanding and elucidating the important interactions between the host gut and its microbiome.

METHODS OF ETBF DETECTION

Currently, simple methods to detect and characterize ETBF colonization or infection in clinical laboratories do not exist. Detection is done through a combination of bacterial isolation and *bft* gene detection or *in-vitro* evaluation of BFT biological activity.

Basic Microbiologic Approaches to Isolating *B. fragilis* Strains

There are a variety of basic microbiologic techniques used to isolate single strains of *B. fragilis* from a variety of different samples. *B. fragilis* is an obligate anaerobe, therefore, it is important to note that care should be taken to decrease the amount of time collected specimens are exposed to the air. Use of anaerobic jars and transport media will help to increase the specimen viability [7].

Isolation From Stool

Stool samples can be collected in two ways: rectal swabs or bulk stool collection. Rectal swabs are streaked on selective media plates and single colonies are isolated and grown separately to test for *B. fragilis* [18]. Bulk stool collection is more effective in characterizing *B. fragilis* as there is a larger amount of material available for use and the samples can be stored at -80°C for long periods of time without affecting *B. fragilis* isolation ability [24]. Samples collected from bulk stool are then plated on Bacteroides Bile Esculin (BBE) selective media, grown anaerobically at 37°C to selectively grow strains from the *B. fragilis* group. *B. fragilis* grown on BBE media will present as raised grey colonies that have hydrolyzed esculin turning the plate black [25]. Furthermore, isolation of *B. fragilis* can be enhanced by anaerobically growing a portion of the stool sample in a non-selective broth media such as Brain Heart Infusion (BHI) prior to plating [24].

Isolation From Blood or Tissue

Tissue samples are also processed for isolation using two methods: broth amplification or tissue homogenization. Broth amplified tissue samples are placed directly in Peptone Yeast Glucose Bile broth (PYGB) following collection. These samples are incubated anaerobically at 37°C and plated on Brucella Blood Agar (BRU) and BBE once turbid. Tissue samples that are processed *via* homogenization are collected in anaerobic transport media, washed in dithiothreitol, and homogenized using a pestle under anaerobic conditions and plated on BRU and BBE following homogenization. Colonies that grow on BRU are then sub-cultured to BBE in order to select for *B. fragilis*. Addition of this extra growth step on BRU non-selective media can increase the isolation rates of *B. fragilis* [4]. In addition to BRU, Brucella Laked Blood Agar with Kanamycin and Vancomycin (LKV) plates have also been used [3]. The strains isolated using these methods are then saved for further down-stream analysis. Isolation of single colonies using these basic microbiological methods are necessary if characterization of individual *B. fragilis* isolates is required. Rapid testing for the presence or absence of *B. fragilis* can be conducted using fecal DNA extraction techniques if strain isolation is not necessary.

BFT Toxin Assay

Early on, BFT biological activity was assessed using lamb ileal loop and reversible ileal tie adult rabbit diarrhea (RITARD) assays [18, 26]. These expensive and labor-intensive methods required the injection of isolated strains into sutured intestinal loops to measure subsequent fluid accumulation as a result of toxin response. *In-vitro* detection methods were then developed using cell culture based HT29/C1 cell assay. This cell assay detects as low as 0.5 pM BFT with 100% specificity when compared to lamb ileal loop assay by microscopically observing the cellular morphological changes caused by BFT [27, 28]. For more than two decades this cell assay has played an important role in assessing *B. fragilis* isolated from clinical samples in order to diagnose ETBF infection and characterize the relative amount of toxin produced by ETBF strains. In the laboratory setting, this assay has been widely used to elucidate the mechanism through which BFT effects intestinal cells [16, 17, 29, 30]. The limitation of this cell assay is the requirement of the cell culture and in most cases, overnight growth of bacterial isolates.

Table 1. PCR primer sets used to characterize *B. fragilis* and BFT gene isotypes.

Primer name	Primer sequence (5' →3')	Reference strain / gene access	Bacterial target	Gene	Reference(s)
Leu-3	CACTTGACTGTTGTAGATAAAGC	CR626927.1, AP006841.1, NC_016776.1	<i>B. fragilis</i>	<i>leuB</i>	[37]
Leu-4	CATCTTCATTGCAGCATTATCC				
Leu-Prb	TGTGCTTGCTTCCAGTCGTCTATG				
Bf904F	GGCGGTCCTCCGGGTTAAA	GU130198; GU130199; GU130200, CR626927	<i>B. fragilis</i>	<i>gyrB</i>	[32, 38]
Bf958R	CACACTTCTGCGGGTCTTTTGT				
Bf923-MGB	FAM-TGGCCGACTGCTC-MGBNFQ				
qHS601F	GTTGTGAAAGTTTTCGGGCTCA	ATCC25285/CR626927	<i>B. fragilis</i>	16S rRNA	[33, 39]
qBAc725R	CAATCGGAGTTCTTCGTGATATCTA				
qHS624MGB	FAM-CGTAATAATTGCAGTTGA-MGB				
F	TCRGGAAAGAAAGCTTGCT	ATCC25285/CR626927	<i>B. fragilis</i>	16S rRNA	[31] ^a
R	CATCCTTTACCGGAATCCT				
Prob	ACACGTATCCAACCTGCCCTTACTCG				
bftF	GGG ACA AGG ATT CTA CCA GCT TTA TA	VPI 13784 (bft-1), 86-5443-2-2 (bft-2)	ETBF	<i>bft</i>	[36, 40]
bftR	ATT CGG CAA TCT CAT TCA TCA TT				
bftProb	CGC AAT GGC GAA TCC ATC AGC TAC A				
bft-1F	GGG ATG TCC TGG TTC A	AB026625	ETBF	<i>bft-1</i>	[36, 40]
bft-1R	AAT TAT CCG TAT GCT CAG CG				
bft-1Prob	CTT CGG ATT TTR AAG CCA GTG GGA TGT C				
bft-2F	CTT AGG CAT ATC TTG GCT TG	AB026626	ETBF	<i>bft-2</i>	[36, 40]
bft-2R	GCG ATT CTA TAC ATG TTC TC				
bft-2Prob	CTT CGG ATT TTR AAG CCA GTG GGA TGT C				
bft-3F	TTT GGG CAT ATC TTG GCT CA	AB026624	ETBF	<i>bft-3</i>	[36, 40]
bft-3R	ATC ATC CGC ATG GTT AGC A				
bft-3Prob	CTT CGG ATT TTR AAG CCA GTG GGA TGT C				

^aPCR primers/probes to detect other members in *B. fragilis* group were also described in this report.

PCR Based Detection

Detection of *B. fragilis*

Polymerase chain reactions (PCR) are an important method in *B. fragilis* and BFT detection due to its quick turnaround time (hours) as well as independence from strain viability and storage conditions. PCR based detection allows for flexibility in collection methods used in gathering samples. A large number of PCR methods for the detection of *B. fragilis* have been characterized and previously reported at length in a review by C.L. Sears in 2009, including conventional, nested, and multiplex PCR [14]. Multiplex PCR reactions are more specific with the ability to differentiate between species of the *B. fragilis* group present from a series of four multiplex PCR reactions [6]. More recent real-time PCR (RT-PCR) protocols targeting 16S rRNA for detection of *B. fragilis* group species with lower detection limits were also reported [31]. PCR based on genes specific to *B. fragilis* have also been reported to achieve higher specificity of detection. Beta-isopropylmalate dehydrogenase (*leuB*), a gene used in leucine biosynthesis, and the

B-subunit of DNA gyrase (*gyB*) are both genes that have been used as PCR targets to detect *B. fragilis* [32 - 34]. These reports compared RT-PCR protocols with conventional PCR and RT-PCR developed based on a set of primers and probes that amplified 16S rRNA regions conserved among *B. fragilis* group species and concluded that RT-PCRs targeting single-copied genes show better specificity. These PCR protocols are capable to rapidly characterize *B. fragilis* group species and have become an important tool for discerning clinical infections and identification of relevant bacterium.

Detection of Enterotoxigenic *B. fragilis* (ETBF)

Conventional PCR assays detecting BFT have been successful in determining the specific isotype of toxin present in culture by attempting to identify the presence of *bft* gene. Detection of the *bft* gene without characterization of the toxin isotype has been described using boiled bacterial DNA obtained from cultured isolates [3]. Multiplex PCR assays able to identify the toxin subtype present in boiled bacterial DNA samples have also been developed [35]. These assays are able to quickly confirm the identity of relevant clinical strains but are only useful for conventionally isolated bacteria.

Real Time PCR Detection of *B. fragilis*

Detection methods using real time or quantitative PCR are especially useful to determine the presence of *B. fragilis* in mixed clinical samples. A series of relevant real-time PCR primer sets for the detection of *B. fragilis* and BFT isotypes are shown in Table 1. Detection of the *bft* gene in DNA extracted directly from stool samples of patients with diarrhea was used in development of a quantitative PCR assay. The developed assay was capable of determining the *bft* gene isotype as well [36]. Development of q-PCR assays capable of directly testing DNA isolated from patient stool is extremely useful in the detection of relevant clinical infections. Other experiments have shown that incubation of stool samples in PYBG broth assists in increasing the sensitivity of detection [24]. A study comparing conventional and real-time PCR detection of *B. fragilis* indicated that conventional PCR reactions were less sensitive, but the overall turn-around time for detection and analysis was similar [37].

FUTURE PERSPECTIVES

Nearly three decades have passed since enterotoxigenic *B. fragilis* was first identified as a pathogen in diarrheal diseases of both human and animals. More recent studies associating ETBF with colorectal cancer highlight the importance of this bacterium in clinical and public health settings [4].

Traditional bacterial culture method will remain essential for isolation and characterization of *B. fragilis* and ETBF. However, current efforts to simplify anaerobic bacterial culture processes using antioxidants can potentially help with decreasing labor time and sensitivity associated with anaerobic culture processing [41].

Improving detection methods is especially useful for the clinical diagnostic laboratory. Development of TaqMan RT-PCR based high-throughput direct detection of ETBF from stool and other clinical samples will minimize labor and time involved while allowing for automated interpretation of results with high sensitivity and specificity. Recent commercialized digital PCR (dPCR) technology may become an important tool in detecting ETBF in fecal and environmental samples as it is capable of offering quantifiable and more accurate detection of ETBF directly in the gut microbiome. Recently, a new culture based system termed culturomics has been described [42]. By using various antibiotics and inhibitors this method allows for the selective growth of bacterial species present in low quantities in gut populations. When it is combined with matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) or 16S rRNA sequencing, this method may provide a way to overcome the drawback of the current metagenomics approach in studying minor populations of the gut microbiome. In the future, this combined system may replace current phenotypic methods in clinical laboratories allowing for rapid identification of microbial species in stool and other samples. ELISAs are very useful tools in being able to detect the presence of pathogens in patient samples. Measurements of human serum samples show that individuals infected with ETBF had increased levels of both IgA and IgG against BFT when compared to controls [3]. A sensitive and specific ELISA assays needs to be optimized and applied to clinical studies as ELISAs will play a crucial role in quick diagnostic tests to characterize ETBF strains in patient samples and allow for a longitudinal investigation of ETBF infections. Current work is being done by this lab to develop a BFT ELISA in order to ultimately develop a rapid test capable of identifying BFT responses in the clinical laboratory.

Within the past two decades there has been tremendous advancements in molecular biology techniques capable of not only detecting clinically relevant information regarding specific pathogens but also in assisting with the determination of specific host-pathogen interactions at a genetic level. The tools outlined in this review are crucial in

assisting in expanding our understanding of these pathogens and their disease relationships. Currently, steps are being taken to increase the sensitivity of many of these detection methods. We hope that further advances in the implementation of molecular biology techniques will allow for a more comprehensive understanding of the role of enterotoxigenic *B. fragilis* in the human gut microbiome.

CONCLUSION

There has been abundant research conducted in an attempt to characterize the role of enterotoxigenic *B. fragilis* in human gut dysbiosis and clinical pathologies. The ability to effectively isolate and characterize these strains from clinical or laboratory specimens is essential for further investigating the important impact these bacterium have on human colon health. In this review we provided a brief overview of the various gross microbiologic techniques used to isolate bacterial colonies from various sample types. Additionally, molecular biology techniques currently available to properly characterize these strains as well as assays to assess biological activity are reviewed. While there are well established techniques suitable for the isolation and characterization of ETBF the implementation of high throughput screening methodologies and immunoassays currently being developed will assist researchers to characterize these species in a more sensitive manner.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

- [1] Polk BF, Kasper DL. *Bacteroides fragilis* subspecies in clinical isolates. *Ann Intern Med* 1977; 86(5): 569-71. [<http://dx.doi.org/10.7326/0003-4819-86-5-569>] [PMID: 322563]
- [2] Goldstein EJ. Anaerobic bacteremia. *Clin Infect Dis* 1996; 23(Suppl. 1): S97-S101. [http://dx.doi.org/10.1093/clinids/23.Supplement_1.S97] [PMID: 8953114]
- [3] Sears CL, Islam S, Saha A, *et al.* Association of enterotoxigenic *Bacteroides fragilis* infection with inflammatory diarrhea. *Clin Infect Dis* 2008; 47(6): 797-803. [<http://dx.doi.org/10.1086/591130>] [PMID: 18680416]
- [4] Boleij A, Hechenbleikner EM, Goodwin AC, *et al.* The *Bacteroides fragilis* toxin gene is prevalent in the colon mucosa of colorectal cancer patients. *Clin Infect Dis* 2015; 60(2): 208-15. [<http://dx.doi.org/10.1093/cid/ciu787>] [PMID: 25305284]
- [5] Sack RB, Myers LL, Almeida-Hill J, *et al.* Enterotoxigenic *Bacteroides fragilis*: epidemiologic studies of its role as a human diarrhoeal pathogen. *J Diarrhoeal Dis Res* 1992; 10(1): 4-9. [PMID: 1619243]
- [6] Liu C, Song Y, McTeague M, Vu AW, Wexler H, Finegold SM. Rapid identification of the species of the *Bacteroides fragilis* group by multiplex PCR assays using group- and species-specific primers. *FEMS Microbiol Lett* 2003; 222(1): 9-16. [[http://dx.doi.org/10.1016/S0378-1097\(03\)00296-9](http://dx.doi.org/10.1016/S0378-1097(03)00296-9)] [PMID: 12757940]
- [7] Wexler HM. *Bacteroides*: the good, the bad, and the nitty-gritty. *Clin Microbiol Rev* 2007; 20(4): 593-621. [<http://dx.doi.org/10.1128/CMR.00008-07>] [PMID: 17934076]
- [8] Salyers AA. *Bacteroides* of the human lower intestinal tract. *Annu Rev Microbiol* 1984; 38: 293-313. [<http://dx.doi.org/10.1146/annurev.mi.38.100184.001453>] [PMID: 6388494]
- [9] Wick EC, Sears CL. *Bacteroides* spp. and diarrhea. *Curr Opin Infect Dis* 2010; 23(5): 470-4. [<http://dx.doi.org/10.1097/QCO.0b013e32833da1eb>] [PMID: 20697287]
- [10] Troy EB, Kasper DL. Beneficial effects of *Bacteroides fragilis* polysaccharides on the immune system. *Front Biosci (Landmark Ed)* 2010; 15: 25-34. [<http://dx.doi.org/10.2741/3603>] [PMID: 20036803]
- [11] Franco AA, Cheng RK, Chung GT, Wu S, Oh HB, Sears CL. Molecular evolution of the pathogenicity island of enterotoxigenic *Bacteroides fragilis* strains. *J Bacteriol* 1999; 181(21): 6623-33. [PMID: 10542162]
- [12] Sears CL. The toxins of *Bacteroides fragilis*. *Toxicon* 2001; 39(11): 1737-46. [[http://dx.doi.org/10.1016/S0041-0101\(01\)00160-X](http://dx.doi.org/10.1016/S0041-0101(01)00160-X)] [PMID: 11595636]
- [13] Wu S, Dreyfus LA, Tzianabos AO, Hayashi C, Sears CL. Diversity of the metalloprotease toxin produced by enterotoxigenic *Bacteroides fragilis*. *Infect Immun* 2002; 70(5): 2463-71.

- [http://dx.doi.org/10.1128/IAI.70.5.2463-2471.2002] [PMID: 11953383]
- [14] Sears CL. Enterotoxigenic *Bacteroides fragilis*: a rogue among symbiotes. *Clin Microbiol Rev* 2009; 22(2): 349-69. [Table of Contents.]. [http://dx.doi.org/10.1128/CMR.00053-08] [PMID: 19366918]
- [15] Wu S, Shin J, Zhang G, Cohen M, Franco A, Sears CL. The *Bacteroides fragilis* toxin binds to a specific intestinal epithelial cell receptor. *Infect Immun* 2006; 74(9): 5382-90. [http://dx.doi.org/10.1128/IAI.00060-06] [PMID: 16926433]
- [16] Wu S, Lim KC, Huang J, Saidi RF, Sears CL. *Bacteroides fragilis* enterotoxin cleaves the zonula adherens protein, E-cadherin. *Proc Natl Acad Sci USA* 1998; 95(25): 14979-84. [http://dx.doi.org/10.1073/pnas.95.25.14979] [PMID: 9844001]
- [17] Wu S, Powell J, Mathioudakis N, Kane S, Fernandez E, Sears CL. *Bacteroides fragilis* enterotoxin induces intestinal epithelial cell secretion of interleukin-8 through mitogen-activated protein kinases and a tyrosine kinase-regulated nuclear factor-kappaB pathway. *Infect Immun* 2004; 72(10): 5832-9. [http://dx.doi.org/10.1128/IAI.72.10.5832-5839.2004] [PMID: 15385484]
- [18] Myers LL, Firehammer BD, Shoop DS, Border MM. *Bacteroides fragilis*: a possible cause of acute diarrheal disease in newborn lambs. *Infect Immun* 1984; 44(2): 241-4. [PMID: 6538870]
- [19] Myers LL, Shoop DS, Firehammer BD, Border MM. Association of enterotoxigenic *Bacteroides fragilis* with diarrheal disease in calves. *J Infect Dis* 1985; 152(6): 1344-7. [http://dx.doi.org/10.1093/infdis/152.6.1344] [PMID: 3905990]
- [20] Rhee KJ, Wu S, Wu X, *et al.* Induction of persistent colitis by a human commensal, enterotoxigenic *Bacteroides fragilis*, in wild-type C57BL/6 mice. *Infect Immun* 2009; 77(4): 1708-18. [http://dx.doi.org/10.1128/IAI.00814-08] [PMID: 19188353]
- [21] Wu S, Rhee KJ, Albesiano E, *et al.* A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. *Nat Med* 2009; 15(9): 1016-22. [http://dx.doi.org/10.1038/nm.2015] [PMID: 19701202]
- [22] Wick EC, Rabizadeh S, Albesiano E, *et al.* Stat3 activation in murine colitis induced by enterotoxigenic *Bacteroides fragilis*. *Inflamm Bowel Dis* 2014; 20(5): 821-34. [http://dx.doi.org/10.1097/MIB.000000000000019] [PMID: 24704822]
- [23] Toprak NU, Yagci A, Gulluoglu BM, *et al.* A possible role of *Bacteroides fragilis* enterotoxin in the aetiology of colorectal cancer. *Clin Microbiol Infect* 2006; 12(8): 782-6. [http://dx.doi.org/10.1111/j.1469-0691.2006.01494.x] [PMID: 16842574]
- [24] Chen LA, Van Meerbeke S, Albesiano E, *et al.* Fecal detection of enterotoxigenic *Bacteroides fragilis*. *Eur J Clin Microbiol Infect Dis* 2015; 34(9): 1871-7. [http://dx.doi.org/10.1007/s10096-015-2425-7] [PMID: 26173688]
- [25] Ramamurthy D, Pazhani GP, Sarkar A, *et al.* Case-control study on the role of enterotoxigenic *Bacteroides fragilis* as a cause of diarrhea among children in Kolkata, India. *PLoS One* 2013; 8(4): e60622. [http://dx.doi.org/10.1371/journal.pone.0060622] [PMID: 23577134]
- [26] Myers LL, Shoop DS, Stackhouse LL, *et al.* Isolation of enterotoxigenic *Bacteroides fragilis* from humans with diarrhea. *J Clin Microbiol* 1987; 25(12): 2330-3. [PMID: 3429625]
- [27] Weikel CS, Grieco FD, Reuben J, Myers LL, Sack RB. Human colonic epithelial cells, HT29/C1, treated with crude *Bacteroides fragilis* enterotoxin dramatically alter their morphology. *Infect Immun* 1992; 60(2): 321-7. [PMID: 1730463]
- [28] Saidi RF, Sears CL. *Bacteroides fragilis* toxin rapidly intoxicates human intestinal epithelial cells (HT29/C1) *in vitro*. *Infect Immun* 1996; 64(12): 5029-34. [PMID: 8945542]
- [29] Wu S, Morin PJ, Maouyo D, Sears CL. *Bacteroides fragilis* enterotoxin induces c-Myc expression and cellular proliferation. *Gastroenterology* 2003; 124(2): 392-400. [http://dx.doi.org/10.1053/gast.2003.50047] [PMID: 12557145]
- [30] Mundy LM, Sears CL. Detection of toxin production by *Bacteroides fragilis*: assay development and screening of extraintestinal clinical isolates. *Clin Infect Dis* 1996; 23(2): 269-76. [http://dx.doi.org/10.1093/clinids/23.2.269] [PMID: 8842262]
- [31] Tong J, Liu C, Summanen P, Xu H, Finegold SM. Application of quantitative real-time PCR for rapid identification of *Bacteroides fragilis* group and related organisms in human wound samples. *Anaerobe* 2011; 17(2): 64-8. [http://dx.doi.org/10.1016/j.anaerobe.2011.03.004] [PMID: 21439390]
- [32] Lee CS, Lee J. Evaluation of new gyrB-based real-time PCR system for the detection of *B. fragilis* as an indicator of human-specific fecal contamination. *J Microbiol Methods* 2010; 82(3): 311-8. [http://dx.doi.org/10.1016/j.mimet.2010.07.012] [PMID: 20643165]

- [33] Lee CS, Marion JW, Lee J. A novel genetic marker for the rapid detection of *Bacteroides fragilis* in recreational water as a human-specific faecal indicator. *J Water Health* 2011; 9(2): 253-64. [http://dx.doi.org/10.2166/wh.2011.120] [PMID: 21942191]
- [34] Miki T, Kuwahara T, Nakayama H, *et al.* Simultaneous detection of *Bacteroides fragilis* group species by leuB-directed PCR. *J Med Invest* 2005; 52(1-2): 101-8. [http://dx.doi.org/10.2152/jmi.52.101] [PMID: 15751280]
- [35] Avila-Campos MJ, Liu C, Song Y, Rowlinson MC, Finegold SM. Determination of bft gene subtypes in *Bacteroides fragilis* clinical isolates. *J Clin Microbiol* 2007; 45(4): 1336-8. [http://dx.doi.org/10.1128/JCM.02108-06] [PMID: 17301286]
- [36] Merino VR, Nakano V, Liu C, Song Y, Finegold SM, Avila-Campos MJ. Quantitative detection of enterotoxigenic *Bacteroides fragilis* subtypes isolated from children with and without diarrhea. *J Clin Microbiol* 2011; 49(1): 416-8. [http://dx.doi.org/10.1128/JCM.01556-10] [PMID: 20980581]
- [37] Papaparaskavas J, Mela V, Houhoula DP, Pantazatou A, Petrikos GL, Tsakris A. Comparative evaluation of conventional and real-time PCR assays for detecting *Bacteroides fragilis* in clinical samples. *J Clin Microbiol* 2013; 51(5): 1593-5. [http://dx.doi.org/10.1128/JCM.00449-13] [PMID: 23447634]
- [38] Ji D, Huang IH, Lai CC, *et al.* Prevalence and characterization of enterotoxigenic *Bacteroides fragilis* and toxigenic *Clostridium difficile* in a Taipei emergency department. *J Microbiol Immunol Infect* 2014; S1684-1182(14)00255-2. [PMID: 25648668]
- [39] Okabe S, Shimazu Y. Persistence of host-specific *Bacteroides-Prevotella* 16S rRNA genetic markers in environmental waters: effects of temperature and salinity. *Appl Microbiol Biotechnol* 2007; 76(4): 935-44. [http://dx.doi.org/10.1007/s00253-007-1048-z] [PMID: 17598108]
- [40] Franco AA, Mundy LM, Trucksis M, Wu S, Kaper JB, Sears CL. Cloning and characterization of the *Bacteroides fragilis* metalloprotease toxin gene. *Infect Immun* 1997; 65(3): 1007-13. [PMID: 9038310]
- [41] La Scola B, Khelaifia S, Lagier JC, Raoult D. Aerobic culture of anaerobic bacteria using antioxidants: a preliminary report. *Eur J Clin Microbiol Infect Dis* 2014; 33(10): 1781-3. [http://dx.doi.org/10.1007/s10096-014-2137-4] [PMID: 24820294]
- [42] Lagier JC, Armougom F, Million M, *et al.* Microbial culturomics: paradigm shift in the human gut microbiome study. *Clin Microbiol Infect* 2012; 18(12): 1185-93. [http://dx.doi.org/10.1111/1469-0691.12023] [PMID: 23033984]

Received: July 13, 2015

Revised: October 20, 2015

Accepted: October 22, 2015

© Fathi and Wu; Licensee *Bentham Open*.

This is an open access article licensed under the terms of the Creative Commons Attribution-Non-Commercial 4.0 International Public License (CC BY-NC 4.0) (<https://creativecommons.org/licenses/by-nc/4.0/legalcode>), which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.