

Effect of Co-Inoculation with Mycorrhiza and Rhizobia on the Nodule Trehalose Content of Different Bean Genotypes

L. Ballesteros-Almanza¹, J. Altamirano-Hernandez¹, J.J. Peña-Cabriales³, G. Santoyo¹, J.M. Sanchez-Yañez¹, E. Valencia-Cantero¹, L. Macias-Rodriguez¹, J. Lopez-Bucio¹, R. Cardenas-Navarro² and R. Farias-Rodriguez^{1,*}

¹Instituto de Investigaciones Químico-Biológicas. Universidad Michoacana de San Nicolas de Hidalgo, Edificio B1 Ciudad Universitaria. Morelia, Michoacan, Mexico

²Instituto de Investigaciones Agropecuarias y Forestales. Universidad Michoacana de San Nicolas de Hidalgo, Km 9.5 Carretera Morelia-Zinapécuaro, Unidad Posta Zootécnica. Tarimbaro, Michoacan, Mexico

³Departamento de Biotecnología y Bioquímica, CINVESTAV Campus Guanajuato. Km. 9.6 Carretera Salamanca-Leon. Irapuato, Guanajuato, Mexico

Abstract: Studies on *Rhizobium*-legume symbiosis show that trehalose content in nodules under drought stress correlates positively with an increase in plant tolerance to this stress. Fewer reports describe trehalose accumulation in mycorrhiza where, in contrast with rhizobia, there is no flux of carbohydrates from the microsymbiont to the plant. However, the trehalose dynamics in the Mycorrhiza-Rhizobium-Legume tripartite symbiosis is unknown. The present study explores the role of this tripartite symbiosis in the trehalose content of nodules grown under contrasting moisture conditions. Three wild genotypes (*P. filiformis*, *P. acutifolis* and *P. vulgaris*) and two commercial genotypes of *Phaseolus vulgaris* (Pinto villa and Flor de Mayo) were used. Co-inoculation treatments were conducted with *Glomus intraradices* and a mixture of seven native rhizobial strains, and trehalose content was determined by GC/MS. The results showed a negative effect of mycorrhizal inoculation on nodule development, as mycorrhized plants showed fewer nodules and lower nodule dry weight compared to plants inoculated only with *Rhizobium*. Mycorrhizal colonization was also higher in plants inoculated only with *Glomus* as compared to plants co-inoculated with both microsymbionts. In regard to trehalose, co-inoculation negatively affects its accumulation in the nodules of each genotype tested. However, the correlation analysis showed a significantly positive correlation between mycorrhizal colonization and nodule trehalose content.

Keywords: Mycorrhiza-Rhizobium co-inoculation, Trehalose, Drought stress.

INTRODUCTION

Common beans represent the most important source of dietary protein in developing countries. However, in most of these countries, its cropping has been relegated to marginal production areas, characterized for their poorly developed soils and low nutrient content. Generally speaking, beans are sown under rainfed systems which expose the plant to variable periods of drought. In this regard, an important effort has been made to produce common bean cultivars tolerant to these challenging conditions. An agronomic practice associated with increased yield in these systems is inoculation with *Rhizobium*. This type of inoculation improves not only the nutritional status of nodulated plants, derived from the biological nitrogen fixation (BNF) process, but recently it has been recognized that this inoculation may also increase drought or osmotic tolerance of nodulated legumes such as acacia, common bean, lotus, and medicago, through the

biosynthesis of trehalose (α -D-glucopyranosyl-[1-1]- α -D-glucopyranoside) [1-4]. The accumulation of trehalose by different organisms (bacteria, yeast, fungi, nematodes, etc.) has been related to survival under different environmental stresses, like osmotic, low pH, high and low temperatures and dehydration [5-7]. In the nodule, disaccharide is synthesized by the bacteroid, and its accumulation depends on the rhizobial strain, legume genotype and environmental conditions [8]. It was reported that under well-watered conditions, there is a basal amount of trehalose in the nodules, having a negative correlation with the BNF, but under drought conditions, nodule trehalose levels increase significantly and may contribute to maintain BNF under these circumstances [2] and to improve the plant tolerance to drought [2, 8]. Under field conditions, it was observed that trehalose is detected in seeds from nodulated plants, with a positive correlation with seed longevity [9].

It has been reported that mycorrhizal inoculation increases the plant's tolerance to drought. The most accepted mechanism for this effect is the improvement of water-use efficiency of the mycorrhized plant [10-13]. Interestingly, it is also known that ecto and endo mycorrhizae synthesize trehalose [14, 15]. Nevertheless, in this case, it was demon-

*Address correspondence to this author at the Instituto de Investigaciones Químico-Biológicas. Universidad Michoacana de San Nicolas de Hidalgo, Edificio B1 Ciudad Universitaria. Morelia, Michoacan, Mexico; Tel: (+52) 443 3265790; Fax: (+52) 443 3265788; E-mail: rodolfoarias44@yahoo.com.mx

strated through the use of ^{14}C that there is no carbohydrate flux from the fungi to the plant [16], and fungi trehalose is mostly localized at the spores where it may serve, as in *Trichoderma* and *Aspergillus*, to increase the survival of the spores [17-18]. In a recent *in vitro* study, it was observed that mycorrhizal roots under osmotic stress mobilized trehalose from the mycorrhizal root to the external mycelia (Farias-Rodriguez, unpublished data), supporting the idea that trehalose biosynthesis in mycorrhizal symbiosis is only localized in fungal tissues and does not contribute to plant stress tolerance as in *Rhizobium*.

In the case of co-inoculation (rhizobia and mycorrhiza) experiments on legumes, most of the reports argue for a synergistic effect of both microsymbionts on plant development [19]. However, in common bean, examples in the literature show a negative effect of mycorrhiza on nodule development [20]. The objective of this work was to study the effect of mycorrhiza inoculation on trehalose accumulation in the nodule.

MATERIALS AND METHODS

Biological Material

The common bean genotypes used here were the wild accessions *Phaseolus filiformis* P1535309, *P. acutifolius* G40142 and *P. vulgaris* P1325687, and the commercial cultivars *P. vulgaris* cv. Pinto Villa and *P. vulgaris* cv. Flor de Mayo. The seeds were donated by Jorge Acosta Gallegos from INIFAP-Celaya.

Glomus intraradices spores were donated by Javier Villegas from IIQB-UMSNH and maintained at -70°C in a 50% glycerol solution. The *Rhizobium* strains used (Fcin1, Fcin2, Fcin3, Fcin4, Fcin5, Fcin6, and Fcin7) were isolated in a previous experiment [9]. Stock cultures were maintained at -70°C in PY medium (in g L^{-1} : yeast extract 3, $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ 1, casein peptone 5), containing 50% (v/v) glycerol.

Inoculation Treatment

Seeds were surface disinfected with 2.5 M NaOCl for 5 min and rinsed with sterile distilled water before use. Three inoculation treatments were imposed as follows: a) *Rhizobium* inoculation was performed by soaking the seeds for 1 h in a mixture of native rhizobial strains prepared as described by Altamirano-Hernandez *et al.* [9]; b) *Glomus* inoculation was performed during sowing by adding to each seed 1 mL of a *Glomus* spore suspension (30 spores mL^{-1}); c) co-inoculation was performed by first inoculating *Rhizobium* as described previously, then 1 mL of a *Glomus* spore suspension was added to each seed during sowing. Control plants were not inoculated, instead they were watered with a Jensen solution (in g L^{-1} : 1 $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$, 0.2 K_2HPO_4 , 0.2 $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.2 NaCl, 0.1 $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$, and 1 mL of mineral solution [2.86 boric acid, 0.22 $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$, 1.81 $\text{MnCl}_2 \cdot 7\text{H}_2\text{O}$, 0.09 K_2MoO_4]) containing 0.05% KNO_3 .

Drought Stress

Plants from different inoculation treatments (previously described) were watered every three days, and once a week a Jensen solution was used. Drought stress was imposed on plants at the flowering stage (R7) by suspending watering for

16 days. At the end of this period, plant total biomass and leaf area were recorded. From the *Rhizobium* and co-inoculation treatments, nodules were harvested by hand and nodule number and dry weight were measured.

Root samples were collected from co-inoculated and *Glomus*-inoculated plants to measure mycorrhizal colonization.

Trehalose Analysis

Nodule trehalose was extracted by grinding the nodules in boiling 80% (v/v) ethanol and evaporated to dryness before resuspension in 1 mL of 80% (v/v) ethanol. Aliquots of 100 μL from each sample were derivatized to obtain the aldonitrile peracetylated forms of the sugars, 25 μg of perseeitol were added as internal standard, and then analyzed by gas chromatography-mass spectrometry (HP6850-HP5973) on an HP-5 MS column (Agilent Technologies, Palo Alto, CA) as described by Macias-Rodriguez *et al.* [21].

RESULTS

Nodulation Parameters

In general terms, it was observed that co-inoculation of *Glomus* and *Rhizobium* in bean plants had a negative effect on nodulation parameters (Figs. 1 and 2). In this sense, the wild accession of *P. vulgaris* P1325687 showed the highest number of nodules under both watering regimes and inoculation treatments. In the well-watered treatment (Fig. 1a), *P. vulgaris* P1325687 presented on average 500 nodules per plant when inoculated only with the rhizobial mixture, but when co-inoculated with *Glomus*, the number of nodules in this bean genotype significantly decreased ($p < 0.05$) to an average of 300 per plant. The commercial *P. vulgaris* genotypes (Flor de Mayo and Pinto Saltillo) inoculated only with *Rhizobium* presented 65 nodules per plant on average, but when co-inoculated with *Glomus*, the number of nodules decreased to 45. *P. filiformis* and *P. acutifolius* showed a low number of nodules, with an average of 12 nodules per plant; under co-inoculation they showed an average of 7 nodules per plant.

When drought was imposed on these bean plants, the number of nodules decreased in the two inoculation treatments when compared with their well-watered counterparts (Fig. 1b). *P. vulgaris* P1325687 had 350 nodules per plant when inoculated with *Rhizobium*, and only 100 when co-inoculated with *Glomus*. Under watering conditions, there was a significant difference among the two commercial bean cultivars. *P. vulgaris* cv. Flor de Mayo presented 45 nodules when inoculated with *Rhizobium* and 35 when the mycorrhiza was co-inoculated. Interestingly, *P. vulgaris* cv. Pinto Saltillo had 10 nodules when inoculated only with *Rhizobium* and 40 when the mycorrhiza was co-inoculated. This was the only case recorded in this work in which the number of nodules increased with *Glomus* inoculation. In contrast, *P. filiformis* and *P. acutifolius* had 10 nodules with rhizobial inoculation and 4 with co-inoculation under this drought treatment.

Variability in the dry weight of nodules among the different bean genotypes was lower as compared to that recorded for the total number of nodules (Figs. 1 and 2). Under

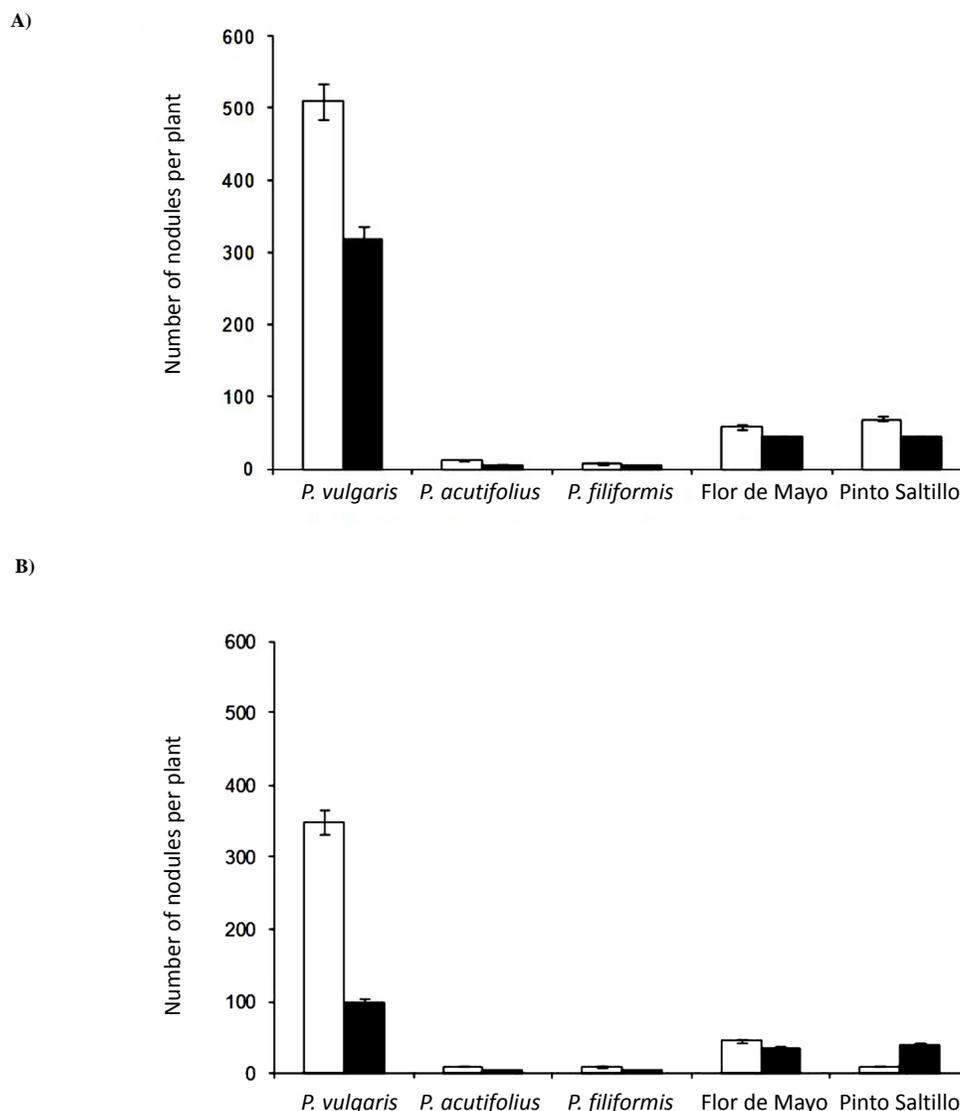


Fig. (1). Number of nodules per plant under **A)** well-watered treatment and **B)** drought stress treatment. White bars represent nodules from plants inoculated only with *Rhizobium*; black bars represent nodules from plants co-inoculated with *Rhizobium* and *Glomus*.

well-watered conditions (Fig. 2a), when plants were inoculated with the rhizobial mixture, *P. acutifolius* showed the largest nodules, with an average dry weight of 0.55 mg nodule⁻¹. *P. vulgaris* P1325687 nodules weighed 0.43 mg on average. The last three genotypes had on average nodules of 0.21 mg. Under the co-inoculation treatment, the weight of nodules significantly ($p < 0.05$) decreased in *P. vulgaris* P1325687 (0.35 mg nodule⁻¹), *P. acutifolius* (0.23 mg nodule⁻¹) and *P. vulgaris* cv. Pinto Saltillo (0.15 mg nodule⁻¹).

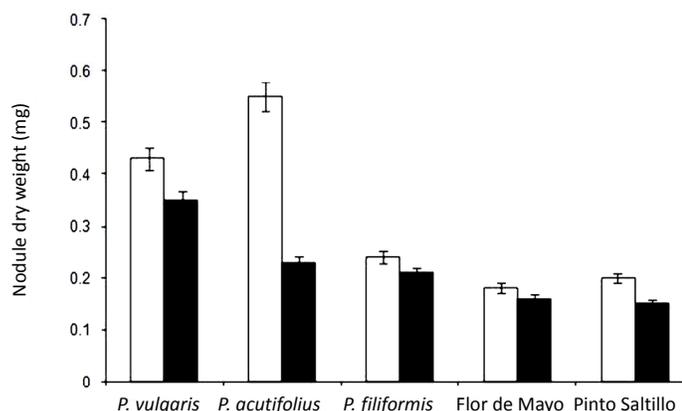
Nodule weight decreased under drought condition (Fig. 2b). As in well-watered plants, *P. acutifolius* had the largest nodules when inoculated only with *Rhizobium*, with an average dry weight of 0.38 mg nodule⁻¹, followed by *P. vulgaris* P1325687 with 0.30 mg nodule⁻¹. The smallest nodules were reordered in *P. vulgaris* cv. Pinto Saltillo with an average of 0.10 mg nodule⁻¹. When these bean genotypes were co-inoculated with *Glomus*, nodule weight decreased significantly ($p < 0.05$) in all genotypes, with a maximum value of

0.19 mg nodule⁻¹ in *P. vulgaris* P1325687, and a minimum of 0.1 mg nodule⁻¹ in *P. acutifolius*. In contrast, *P. vulgaris* cv. Pinto Saltillo nodules showed significant ($p < 0.05$) increases, with 0.16 mg nodule⁻¹.

Nodule Trehalose Content

The effect of mycorrhizal co-inoculation on nodule trehalose content is shown in Table 1. Mycorrhizal co-inoculation caused a decrease in trehalose content of nodules. Under well-watered condition, in plants inoculated only with *Rhizobium*, the average trehalose content in the nodules was 36 mg g⁻¹, the highest value corresponded to *P. filiformis* (50 mg g⁻¹), and the lowest to *P. vulgaris* cv. Flor de Mayo (20 mg g⁻¹). In the co-inoculation treatment, the average nodule trehalose content of these five genotypes significantly ($p < 0.05$) decreased to 16 mg g⁻¹, the maximum value of 25 mg g⁻¹ corresponding to *P. filiformis* and the minimum of 10 mg g⁻¹ to *P. acutifolius* and *P. vulgaris* cv. Flor de Mayo.

A)



B)

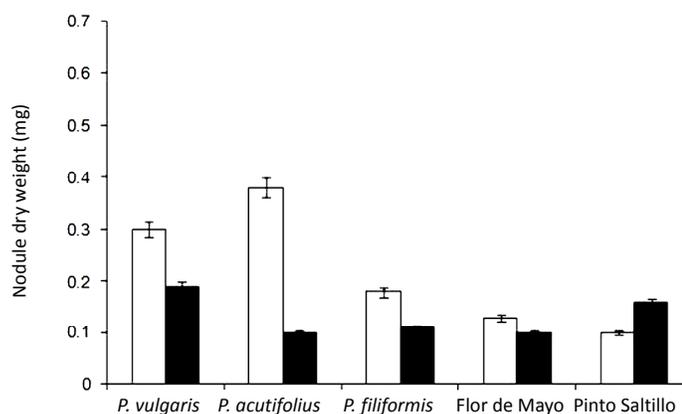


Fig. (2). Nodule dry weight under **A)** well-watered treatment and **B)** drought stress treatment. White bars represent nodules from plants inoculated only with *Rhizobium*; black bars represent nodules from plants co-inoculated with *Rhizobium* and *Glomus*.

Table 1. Effect of Mycorrhizal Inoculation and Drought Stress on Nodule Trehalose Content

	Nodule Trehalose Content (mg g ⁻¹)			
	Well-Watered		Drought Stressed	
	N	N+M	N	N+M
<i>P. filiformis</i>	50	12	90	30
<i>P. acutifolius</i>	30	10	70	30
<i>P. vulgaris</i>	40	25	72	40
<i>P. vulgaris</i> cv. Flor de Mayo	20	10	45	18
<i>P. vulgaris</i> cv. Pinto Saltillo	40	22	60	35

N: inoculated with *Rhizobium*

N+M: inoculated with *Rhizobium* and *Glomus*.

Under drought stress, trehalose content of nodules significantly ($p < 0.05$) increased in both inoculation treatments in comparison to their well-watered counterparts. The average nodule trehalose content for the five genotypes with *Rhizobium* inoculation in this water regime was of 67 mg g⁻¹. The highest trehalose content was recorded in nodules of *P. vulgaris* P1325687 (90 mg g⁻¹), and the lowest

in *P. vulgaris* cv. Flor de Mayo (45 mg g⁻¹). In the co-inoculation treatment, the trehalose content of nodules significantly ($p < 0.05$) decreases in all genotypes, with an average value of 31 mg g⁻¹. The maximum trehalose value corresponded to *P. filiformis* (40 mg g⁻¹) and the minimum to *P. vulgaris* cv. Flor de Mayo (18 mg g⁻¹).

Mycorrhizal Colonization

To better understand the negative effect of *Glomus* inoculation on the nodule parameters described previously, mycorrhizal colonization was recorded and the results are shown in Fig. (3). Interestingly, mycorrhizal colonization was higher when the plants were only inoculated with *Glomus* in all genotypes evaluated, and decreased when the co-inoculation *Glomus-Rhizobium* was performed. Under well-watered conditions, mycorrhizal colonization among the five evaluated bean genotypes was quite uniform, with an average colonization of 22%. The maximum value was recorded for *P. vulgaris* cv. Pinto Saltillo (27%) and the minimum for *P. vulgaris* cv. Flor de Mayo (15%). When co-inoculated with *Rhizobium*, mycorrhizal colonization decreased to an average of 15%. The highest value was 18% for *P. filiformis* and *P. vulgaris* P1325687, and the lowest value was 10% for *P. vulgaris* cv. Flor de Mayo.

Under drought stress it was observed that, as opposed to nodulation, mycorrhizal colonization increased, with an average value of 37%. *P. filiformis* showed the highest colonization with 50%, while *P. vulgaris* cv. Flor de Mayo showed the lowest with 10%. In the co-inoculation treatment, *Glomus* colonization significantly ($p < 0.05$) decreased, as occurred under well-watered conditions, with an average of 21%, the maximum value being for *P. filiformis* (35%) and the minimum for *P. vulgaris* cv. Flor de Mayo (5%).

Correlation Between Nodulation and Mycorrhization

When a correlation analysis was performed to evaluate the effect of mycorrhiza on nodule parameters, it was interesting to observe that under well watered conditions, comparing data from the five bean genotypes, the correlation indices showed a positive relationship between *Glomus* colonization and nodulation (although it wasn't statistically

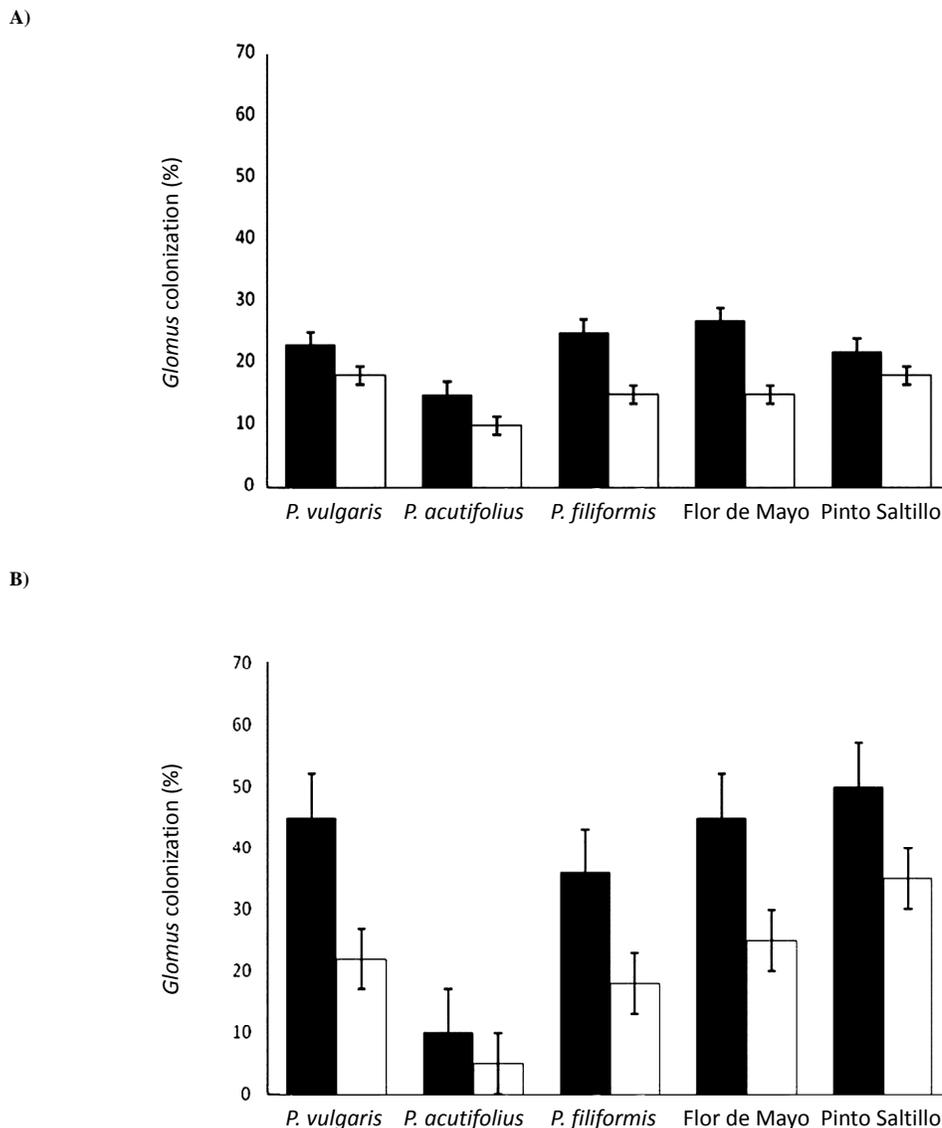


Fig. (3). Percentage of *Glomus* colonization under: A) well-watered treatment and B) drought stress treatment. White bars represent nodules from plants inoculated only with *Glomus*; black bars represent nodules from plants co-inoculated with *Rhizobium* and *Glomus*.

significant). This would indicate that bean genotypes that have the highest mycorrhizal colonization showed the highest number of nodules, nodule dry weight and nodule trehalose content (Fig. 4a), regardless of the fact that individually, in each genotype mycorrhizae decreased the values of these parameters. The correlation index values for *Glomus* colonization were $r=0.38$ for nodule number, $r=0.63$ for nodule dry weight, and $r=0.48$ for trehalose content. However, under drought conditions, some changes were observed in these correlations indices (Fig. 4b). The number of nodules had a negative correlation with mycorrhization ($r=0.15$). Even though the correlation index observed with nodule dry weight decreased, a positive correlation was still observed ($r=0.26$). Interestingly, trehalose correlation index increased to a significant value of 0.98.

Effect of Co-Inoculation on Bean Biomass and Leaf Area

Plant leaf area with the different inoculation treatments under both water regimes is shown in Table 2. Under well-watered conditions, the response of the five bean genotypes to the different inoculation treatments was very heterogeneous. Nevertheless, control plants that were chemically fertilized showed the greatest leaf area (149 cm^2), followed by the plants co-inoculated with *Rhizobium* and *Glomus* (138

cm^2). Plants inoculated with *Rhizobium* had an average leaf area of 118 cm^2 , and the minimum recorded value corresponded to plants inoculated with *Glomus* (103 cm^2). Also, considering only the inoculated treatments, it was observed that in *P. filiformis* and *P. acutifolius* plants, co-inoculation resulted in a negative effect by reducing the leaf area when compared to the maximum area obtained under inoculation, which corresponded to *Rhizobium* inoculation. On the other hand, co-inoculation had a positive effect on *P. vulgaris* P1325687 and *P. vulgaris* cv. Flor de Mayo genotypes, which showed the greatest leaf area under this treatment in comparison to single inoculations.

Under drought stress, bean plants inoculated with *Glomus* presented the greatest leaf area (96 cm^2), followed by plants inoculated with *Rhizobium* (90 cm^2). Leaf area in fertilized control plants decreased to 85 cm^2 , and the smallest leaf area was recorded in co-inoculated plants (84 cm^2). Again, considering only inoculation treatments, it was observed that in *P. acutifolius* and *P. vulgaris* P1325687, the maximum leaf area was obtained under *Glomus* inoculation, and co-inoculation negatively affected this parameter. On the other hand, *P. acutifolius* and *P. vulgaris* cv. Flor de Mayo showed greater leaf area with *Rhizobium* inoculation, being negatively affected by co-inoculation. *P. vulgaris* cv. Pinto

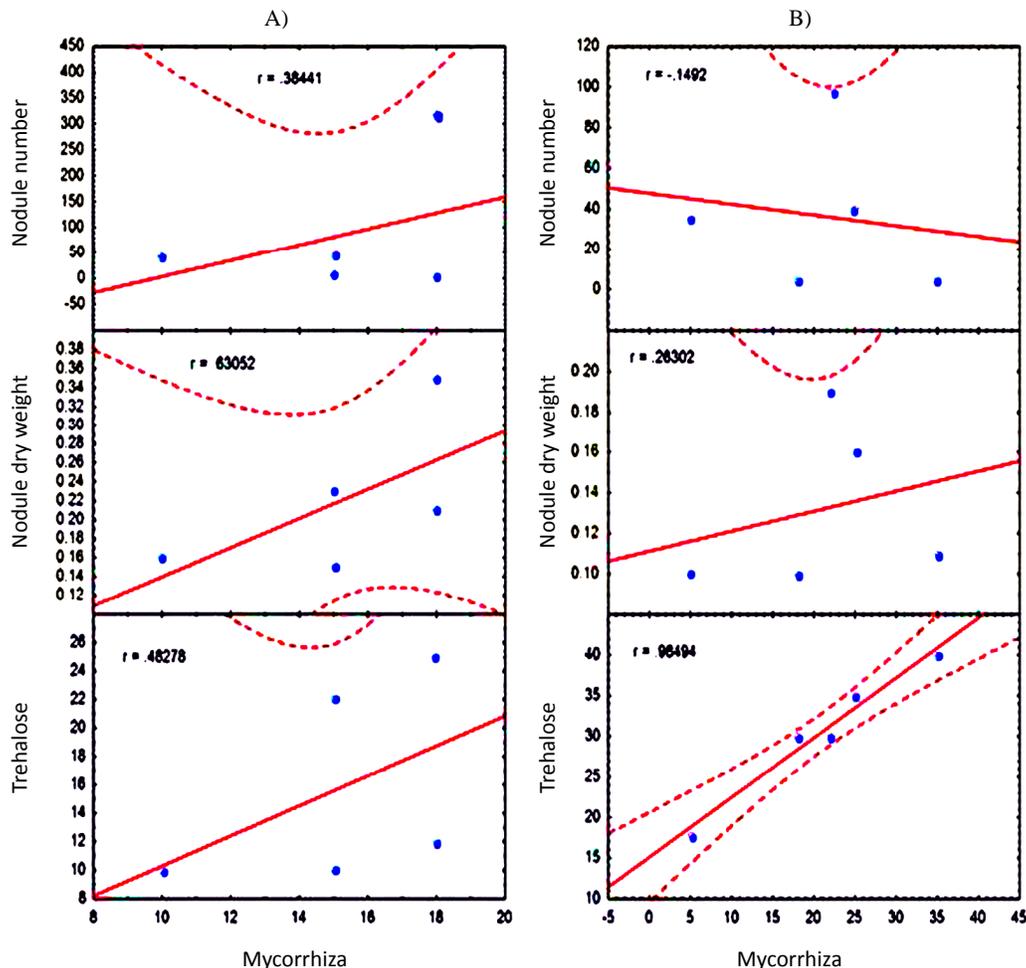


Fig. (4). Correlation indexes between mycorrhiza colonization and nodule parameters; A) well-watered treatment and B) drought stress treatment. Dashed lines represent confidence interval at $p < 0.05$.

Table 2. Effect of Rhizobial and Mycorrhizal Inoculation on Leaf Area of *Phaseolus* Under two Watering Regimes

	Leaf Area (cm ²)							
	Well-Watered				Drought Stressed			
	N	N+M	M	C	N	N+M	M	C
<i>P. filiformis</i>	34a	32a	24a	28a	38a	18b	15b	20b
<i>P. acutifolius</i>	180a	160a	55b	305c	100b	94ab	210b	100b
<i>P. vulgaris</i>	65a	110bc	91b	81b	40a	53ab	66b	66b
<i>P. vulgaris</i> cv. Flor de Mayo	160a	210b	162a	210b	155b	106a	90a	152b
<i>P. vulgaris</i> cv. Pinto Saltillo	152a	180a	185a	120a	115a	149b	100a	86a

N: inoculated with *Rhizobium*.

N+M: inoculated with *Rhizobium* and *Glomus*.

M: inoculated with *Glomus*.

C: chemically fertilized control plants.

Different letters in the same line represent significant differences at $p < 0.05$, under the same irrigation treatment.

Table 3. Effect of Rhizobial and Mycorrhizal Inoculation on Total Biomass of *Phaseolus* Under two Watering Regimes

	Total Biomass (mg plant ⁻¹)							
	Well-Watered				Drought Stressed			
	N	N+M	M	C	N	N+M	M	C
<i>P. filiformis</i>	1.38a	.72b	0.72b	0.6a	.52a	.31b	.37b	.28b
<i>P. acutifolius</i>	2.0a	1.6a	1.7a	2.0a	1.45a	1.0b	0.78b	0.6b
<i>P. vulgaris</i>	1.6a	1.6a	.96a	1.2a	1.2a	.45b	.65b	.35b
<i>P. vulgaris</i> cv Flor de Mayo	2.4a	1.5b	2.0a	1.4b	1.85a	1.15b	1.31b	0.9b
<i>P. vulgaris</i> cv Pinto Saltillo	2.7a	1.45b	2.1a	0.5a	2.0a	1.0b	1.8a	0.9a

N: inoculated with *Rhizobium*.

N+M: inoculated with *Rhizobium* and *Glomus*.

M: inoculated with *Glomus*.

C: chemically fertilized control plants.

Different letters in the same line represent significant differences at $p < 0.05$, under the same irrigation treatment.

Saltillo represented the only case in which a positive effect of co-inoculation was observed, with maximum leaf area.

The effect of inoculation on total biomass is shown in Table 3. In this case, the *Rhizobium* inoculation produced the highest biomass of all five genotypes tested, both under well-watered and drought stressed plants. The total biomass for *Rhizobium* inoculated plants under well-watered conditions was 2 mg plant⁻¹, followed by plants inoculated with *Glomus*, with a biomass of 1.5 mg plant⁻¹, and of the inoculated plants, those that were co-inoculated showed the lower biomass (1.4 mg plant⁻¹). Plants under drought stress inoculated with *Rhizobium* showed a biomass of 1.4 mg plant⁻¹; plants inoculated with *Glomus* showed a biomass of 1 mg plant⁻¹, and those co-inoculated showed 0.7 mg plant⁻¹.

Considering the plant biomass obtained under well-watered conditions as 100% of growth for the different inoculation treatments, inhibition caused by drought was re-

corded and the results are shown in Fig. (5). Although there was no statistical significance ($p < 0.05$) among inoculation treatments, the bean plants less affected by drought were the chemically fertilized ones with an average of 30% biomass reduction when compared to their well-water counterpart.

Among the inoculated treatments, plants inoculated with *Rhizobium* were the least affected by drought (33% biomass reduction), followed by plants inoculated with *Glomus* (37% biomass reduction), and the plants most sensitive to drought were the ones co-inoculated, with an average biomass reduction of 44%.

DISCUSSION

The biosynthesis of trehalose has been correlated with tolerance to different types of stress. Although this carbohydrate is widespread in nature, it is considered to be absent in superior vascular plants in the past decade. Several efforts

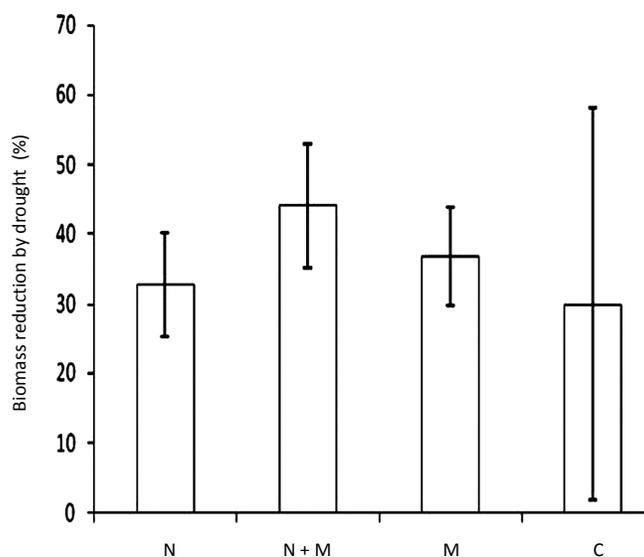


Fig. (5). Percentage of biomass reduction caused by drought in comparison to well-watered plants among the different inoculation treatments. N: Plants inoculated with rhizobia. N+M: Plants co-inoculated with *Glomus* and rhizobia. M: Plants inoculated with *Glomus*. C: Non-inoculated control plants.

have been made to obtain transgenic plants that express the microbial genes for the synthesis of trehalose, with the aim of increasing plants' tolerance to dehydration [22, 23]. Up until today, the presence of trehalose genes in several plants has been reported [24-26], but the carbohydrate has been rarely detected [27], and assays on transgenic plants continue [28-31]. On the other hand, when plants are in symbiosis with different microorganisms, trehalose is commonly detected. The role of trehalose in these interactions has been mostly studied in the *Rhizobium*-legume symbiosis and less work has been conducted in regard to fungi-plant interaction.

Today, there are no reports in the literature that have considered the effect of co-inoculation with bacteria and fungi on plant trehalose accumulation. In this sense, studies on co-inoculation of *Rhizobium* and mycorrhiza on legumes have focused on mycorrhiza performance and its effect on plant nutrient uptake and, only in a few cases, on plant drought tolerance. For example, in faba bean (*Vicia faba*) it has been reported that mycorrhizal inoculation increased nodule number when compared to non-mycorrhized plants, and this positive effect on nodulation was also observed under drought conditions, but when plant drought tolerance was discussed, only the mycorrhizal parameters were considered [32].

There are scarce reports on common bean, and these reports point to a negative effect of mycorrhiza on nodule development and nitrogen fixation [20, 33]. Therefore, considering all the reports that establish that trehalose accumulation in legume nodules is related to increased plant drought stress, the aim of the present work was to analyze the effect of mycorrhizal colonization on nodulated common bean based on parameters related to drought tolerance.

This work is in agreement with previous reports on common bean, which support the idea that co-inoculation has a negative effect on the microsymbionts that are involved in this process [33], since nodulation and mycorrhizal

colonization were higher when inoculated independently, and both parameters decreased with co-inoculation. Of the three wild beans used in this work, *P. filiformis* and *P. acutifolius* showed the lowest number of nodules in all treatments. This could be attributed to the origin of the rhizobial strains, which were isolated from a field devoted to common bean production and may not be the most adequate strains for these two genotypes. On the contrary, there was no difference in mycorrhizal colonization between these three genotypes under well-watered conditions. *P. filiformis* showed high mycorrhizal colonization under drought conditions. When co-inoculation was assayed, both mycorrhizal colonization and nodulation decreased in all bean genotypes tested. This behavior was probably due to a competition for plant photosynthates between both microorganisms, imposed by the experimental conditions used at this work. Interestingly, when the data from the five bean genotypes were combined to compute the correlation indexes, the opposite correlations were observed

Under well-watered conditions, nodule number and dry weight correlate positively with mycorrhizal colonization. In other words, among the different bean genotypes, the higher the mycorrhization the higher the nodulation; however, as mentioned earlier, this type of co-inoculation reduced the value of these two parameters when compared to plants inoculated only with *Rhizobium*. In drought stressed plants, correlation indexes between nodulation and mycorrhizal colonization parameters were negative, indicating that nodulation is more sensitive to dehydration than the mycorrhiza. Also, as mycorrhiza is more widely distributed in the root, there is the possibility that this fungus could better exploit the limited photosynthates available in this condition.

In regard to trehalose accumulation, in co-inoculation treatments there was a decrease in its nodule content when compared with plants inoculated only with rhizobia. It was observed that under both watering regimes, the correlation index was positive, and under drought it became highly significant ($r=0.98$). Up until today, the precise biochemical and

molecular control of trehalose biosynthesis in the nodule is not understood. Although some data suggest that nodule trehalose could be mobilized to the plant, there is no conclusive evidence. Therefore, if there is a trehalose release from the nodule under drought to the plant tissue, it would be possible, under this circumstance, for the mycorrhiza to utilize it and export the trehalose to the external mycelia and spores, and this fungal dependency on nodule trehalose may provoke this high correlation index. The mobilization of trehalose from the mycorrhizal roots to the external mycelia has been previously observed in an experiment using as a study model transformed carrot roots, which once mycorrhized, were submitted to osmotic stress; it was observed that under these conditions, there was a higher trehalose content in the external mycelia as compared to the non-stressed controls (Farias-Rodriguez, unpublished data). As this work focused on reproducing the soil environment under field conditions, it's very difficult to understand the basic mechanisms underlying this observation; in order to elucidate this relationship, future work should be conducted with the use of *in vitro* systems, that will allow a better understanding of the phenomena reported here.

When considering biomass production as an estimate of drought tolerance, it was observed once again that plants inoculated only with rhizobia or mycorrhiza were less affected by drought, whereas when co-inoculated, a higher decrease in biomass production was observed as compared to the plants' well-watered counterparts. In conclusion, the results presented in this work support the idea that the use of rhizobial or mycorrhizal inoculation separately would result in better bean drought tolerance than the co-inoculation treatment.

REFERENCES

- [1] Räsänen LA, Saijets S, Jokinen K, Lindström K. Evaluation of the roles of two compatible solutes, glycine betaine and trehalose, for the *Acacia senegal*-*Sinorhizobium* symbiosis exposed to drought stress. *Plant Soil* 2004; 260: 237-51.
- [2] Jimenez-Zacarias JJ, Altamirano-Hernandez J, Peña-Cabriaes JJ. Nitrogenase activity and trehalose content of nodules of drought-stressed common beans infected with effective (Fix+) and ineffective (Fix-) rhizobia. *Soil Biol Biochem* 2004; 36: 1975-81.
- [3] Lopez M, Herrera-Cervera JA, Lluch C, Tejera NA. Trehalose metabolism in root nodules of the model legume *Lotus japonicus* in response to salt stress. *Physiol Plant* 2006; 128: 701-9.
- [4] Lopez M, Tejera NA, Iribarne C, Lluch C, Herrera-Cervera JA. Trehalose and trehalase in root nodules of *Medicago truncatula* and *Phaseolus vulgaris* in response to salt stress. *Physiol Plant* 2008; 134: 575-82.
- [5] Doehlemann G, Berndt P, Hahn M. Trehalose metabolism is important for heat stress tolerance and spore germination of *Botrytis cinerea*. *Microbiology* 2006; 152: 2625-34.
- [6] McIntyre HJ, Davies H, Hore TA, Miller SH, Dufour JP, Ronson CW. Trehalose biosynthesis in *Rhizobium leguminosarum* bv. *trifolii* and its role in desiccation tolerance. *Appl Environ Microbiol* 2007; 73: 3984-92.
- [7] Mahmud SA, Nagahisa K, Hirasawa T, Yoshikawa K, Ashitani K, Shimizu H. Effect of trehalose accumulation on response to saline stress in *Saccharomyces cerevisiae*. *Yeast* 2009; 26: 17-30.
- [8] Farias-Rodriguez R, Mellor RB, Arias C, Peña-Cabriaes JJ. The accumulation of trehalose in nodules of several cultivars of common bean (*Phaseolus vulgaris*) and its correlation with resistance to drought stress. *Physiol Plant* 1998; 102: 352-9.
- [9] Altamirano-Hernandez J, Lopez MG, Acosta-Gallegos JA, Farias-Rodriguez R, Peña-Cabriaes JJ. Influence of soluble sugars on seed quality in nodulated common bean (*Phaseolus vulgaris* L.): The case of trehalose. *Crop Sci* 2007; 47: 1193-205.
- [10] Ahmad Khan I, Ahmad S, Mirza S. Yield and water use efficiency (WUE) of *Avena sativa* as influenced by vesicular arbuscular mycorrhizae (VAM). *Asian J Plant Sci* 2003; 2: 371-3.
- [11] Bolandnazar S, Aliasgarzad N, Neishabury MR, Chaparzadeh N. Mycorrhizal colonization improves onion (*Allium cepa* L.) yield and water use efficiency under water deficit condition. *Scientia Hort* 2007; 114: 11-15.
- [12] Farahani A, Lebaschi H, Hussein M, Hussein SA, Reza VA, Jahanfar D. Effects of arbuscular mycorrhizal fungi, different levels of phosphorus and drought stress on water use efficiency, relative water content and proline accumulation rate of Coriander (*Coriandrum sativum* L.). *J Med Plants Res* 2008; 2: 125-31.
- [13] Maiquetia M, Caceres A, Herrera A. Mycorrhization and phosphorus nutrition affect water relations and CAM induction by drought in seedlings of *Clusia minor*. *Annals Botany* 2009; 103:525-32.
- [14] Becard G, Doner LW, Rolin DB, Douds DD, Pfeffer PE. Identification and quantification of trehalose in vesicular-arbuscular mycorrhizal fungi by *in vivo* ¹³C NMR and HPLC analyses. *New Phytol* 1991; 118: 547-52.
- [15] Fajardo Lopez M, Männer P, Willmann A, Hampp R, Nehls U. Increased trehalose biosynthesis in Hartig net hyphae of ectomycorrhizas. *New Phytol* 2007; 174: 389-98.
- [16] Pfeffer PE, Douds DD, Bücking H, Schwartz DP, Shachar-Hill Y. The fungus does not transfer carbon to or between roots in an arbuscular mycorrhizal symbiosis. *New Phytol* 2004; 163: 617-27.
- [17] Pedreschi F, Aguilera JM. Viability of dry *Trichoderma harzianum* spores under storage. *Bioprocess Eng* 1997; 17: 177-83.
- [18] Fillinger S, Chaverche MK, van Dijk P, et al. Trehalose is required for the acquisition of tolerance to a variety of stresses in the filamentous fungus *Aspergillus nidulans*. *Microbiology* 2001; 147: 1851-62.
- [19] Kawai Y, Yamamoto Y. Increase in the formation and nitrogen fixation of soybean nodules by vesicular-arbuscular mycorrhiza. *Plant Cell Physiol* 1986; 27: 399-405.
- [20] Antunes PM, de Varennes A, Zhang T, Goss MJ. The tripartite symbiosis formed by indigenous arbuscular mycorrhizal fungi, *Bradyrhizobium japonicum* and soya bean under field conditions. *J Agro Crop Sci* 2006; 192:373-8.
- [21] Macias-Rodriguez L, Quero E, Lopez MG. Carbohydrate differences in strawberry crowns and fruit (*Fragaria × ananassa*) during plant development. *J Agric Food Chem* 2002; 50: 3317-21.
- [22] Holmström KO, Mäntylä E, Welin B, Mandal A, Palva ET, Tunnela OE, Londesborough J. Drought tolerance in tobacco. *Nature* 1996; 379: 683-4.
- [23] Goddijn O, Verwoerd TC, Voogd E, et al. Inhibition of trehalase activity enhances trehalose accumulation in transgenic plants. *Plant Physiol* 1997; 113: 181-90.
- [24] Wang YJ, Hao YJ, Zhang ZG, Chen T, Zhang JS, Chen SY. Isolation of trehalose-6-phosphate phosphatase gene from tobacco and its functional analysis in yeast cells. *J Plant Physiol* 2005; 162: 215-23.
- [25] Garg AK, Kim JK, Ranwala A, Wu R. Accumulation of trehalose in transgenic *indica* rice using bifunctional fusion enzyme of trehalose-6-phosphate synthase and trehalose-6-phosphate phosphatase of *Escherichia coli*. *Rice Genet Newsl* 2009; 18: 97-89.
- [26] NCBI. Available from: <http://www.ncbi.nlm.nih.gov/sites/entrez/>. [Accessed on May 6, 2009].
- [27] Goddijn O, Smeekens S. Sensing trehalose biosynthesis in plants. *Plant J* 1998; 14: 143-6.
- [28] Penna S. Building stress tolerance through over-producing trehalose in transgenic plants. *Trends Plant Sci* 2003; 8: 355-7.
- [29] Karim S, Aronsson H, Ericson H, et al. Improved drought tolerance without undesired side effects in transgenic plants producing trehalose. *Plant Mol Biol* 2007; 64: 371-86.
- [30] Almeida AM, Cardoso LA, Santos DM, Torne JM, Feveireiro PS. Trehalose and its applications in plant biotechnology. *In Vitro Cell Dev Biol Plant* 2007; 43: 167-77.
- [31] Iordachescu M, Imai R. Trehalose biosynthesis in response to abiotic stresses. *J Integrative Plant Biol* 2008; 50: 1223-9.

[32] Abdel-Fattah GM, Fatma, Migahed FF, Ibrahim AH. Interactive effects of endomycorrhizal fungus *Glomus etunicatum* and phosphorous fertilization on growth and metabolic activities of broad beans plants under drought stress conditions. *Pak J Biol Sci* 2002; 5: 835-41.

[33] Bethlenfalvay GJ, Pacovsky RS, Bayne HG, Stafford AE. Interactions between nitrogen fixation, mycorrhizal colonization, and host-plant growth in the *Phaseolus-Rhizobium-Glomus* symbiosis. *Plant Physiol* 1982; 70: 446-50.

Received: May 31, 2010

Revised: June 17, 2010

Accepted: June 25, 2010

© Ballesteros-Almanza *et al.*; Licensee *Bentham Open*.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.